

A DESCRIPTION OF OPEN TOP CHAMBER SYSTEM FOR OIL PALM CARBON DIOXIDE ENRICHMENT

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ABSTRACT

Open Top Chamber (OTC) is one of the techniques to study plant responses to elevated carbon dioxide (CO₂) concentrations. A study on the effects of CO₂ enrichment on oil palm growth, physiology and yield using OTC method was conducted at the Malaysian Palm Oil Board (MPOB) Research Station in Kluang, Johor, Malaysia. Oil palm clones P164 were cultivated in four OTC each with 9 m diameter and 10 m high. The OTC consists of a concrete structure with transparent multi-wall sheets. It has a truncated top for air aversion and to prevent CO₂ dilution within the chamber by outside air. The CO₂ gas was injected from nozzles mounted on the inner wall. The OTC was able to maintain a higher CO₂ level inside the OTC than ambient level. Air temperature inside the OTC was slightly higher than ambient, while relative humidity and photosynthetically active radiation (PAR) inside the OTC were both slightly lower than ambient. A comparison of parameters measured inside the OTC and ambient provides a measure of the chamber effects. However, it could have either positive or negative effect on plant growth. The OTC method has some limitations and is only suitable for studying individual plant physiological responses to elevated CO₂.

Keywords: Open Top Chamber, carbon dioxide enrichment, oil palm physiology, oil palm growth.

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INTRODUCTION

Carbon dioxide (CO₂) is the primary greenhouse gas that has been increasing in the atmosphere through human activities since the industrial revolution era. The contributors are burning fossil fuel (oil, coal and natural gas) for energy in machinery and transportation, open burning as well as deforestation (McCracken, 2008). In general, CO₂ occurs naturally as part of the carbon cycle among the atmosphere, oceans, soil, plants and animals (EPA, 2017). Human activities put in more CO₂ to

the atmosphere and influence the forest as a natural sink and have modified the natural carbon cycle process (Pidwirny, 2006).

Increases in atmospheric CO₂ and other infrared-absorbing greenhouse gasses contribute to a warmer climate in which surface air temperature rises to 1.5°C to 5.5°C (Adams *et al.*, 1990). An increase in global temperatures can cause other changes, including rainfall patterns (Baker and Allen, 1994), melting snow or ice, rising sea level and increasing sea acidity (Barnett *et al.*, 2005). Since 1970 to 2004, CO₂ annual emission increased by about 80% and the projection to continue to increase as much as 500 to 1000 ppm by the year 2100 (IPCC, 2007).

Rising CO₂ levels markedly affect plant growth, physiology, and chemistry (Ziska, 2008). In plant metabolism, CO₂ breaks down into smaller carbon molecules chemically. The products are used in chemical energy and to build plant structure.

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Photosynthesis process assimilates CO₂ and turns it into organic molecules that produce about 96% of the total dry mass of a plant (Marschner, 1995). The main components of photosynthesis process are CO₂, water and light energy, thus increasing CO₂ availability which may affect plant growth and photosynthesis.

The CO₂ fertilising effect of elevated CO₂ may give impact to crop productivity and agro-ecosystems. Under higher CO₂ concentration, the photosynthetic pathway plays an important role that influence plant growth and yield. Different plant species show a different response to elevated CO₂ levels. Plants with C₃ photosynthesis will respond markedly to increasing CO₂ concentrations. Plants with C₄ photosynthesis will show little response to rising atmospheric CO₂ because they have a mechanism to increase the concentration of CO₂ in leaves that causes CO₂ saturation of photosynthesis at current ambient concentrations (Allen and Prasad, 2004).

Being a C₃ plant, oil palm growth and yield are expected to be improved under high CO₂ levels through increased photosynthetic rates, reduced photorespiration, and increased water use efficiency. The previous experiment on oil palm seedlings had shown that CO₂ at 800 ppm increased the seedling biomass by about twice of control (Ibrahim *et al.*, 2010). However, the increasing temperature, poor soil nutrient, and inadequate water supply could limit the advantage of high CO₂ on oil palm growth, fresh fruit bunch production or oil quality. Therefore, increasing amount of CO₂ levels could have both detrimental and beneficial effects on oil palm crop.

Open Top Chamber (OTC) is a method used to study the effects of elevated CO₂ and other atmospheric gases on vegetation. It is a plastic enclosure, with an open top, constructed of metal frame covered by panels of polyvinyl chloride plastic film (Machacova, 2010). A truncated top or frustum averts air and prevents CO₂ dilution within the chamber by outside air. Air is pulled into the bottom of the chamber, enriched with CO₂, and then blown through the open top of the chamber (Figure 1).

OPEN TOP CHAMBER STRUCTURE

The OTC facility for CO₂ enrichment on oil palm is located at the Malaysian Palm Oil Board (MPOB) Research Station in Kluang, Johor, Malaysia. The OTC dimension is 9 m diameter and 10 m high, suitable for growing a single oil palm until maturity (Figure 2). The OTC consists of a concrete structure with aluminum frames to hold the 6 mm thick transparent multi-wall polycarbonate sheet with a direct solar transmission of 76% (Sabic Innovative Plastics, USA). It has a truncated top for air aversion

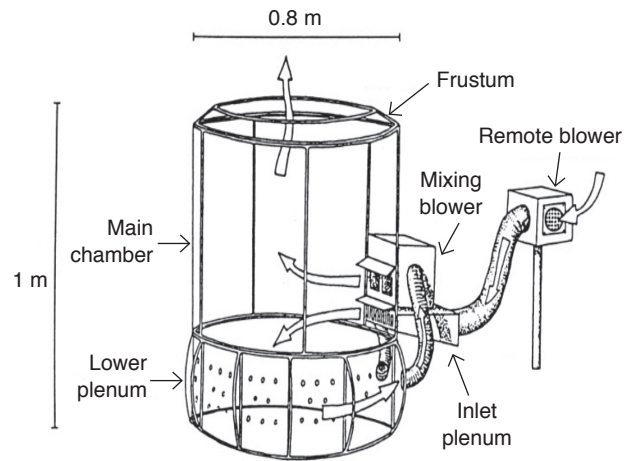


Figure 1. An Open Top Chamber design for salt marsh vegetation by Drake *et al.* (1989).



Figure 2. An Open Top Chamber (10 m high x 9 m diameter) at MPOB Kluang Station, Johor, Malaysia.

and to prevent CO₂ dilution within the chamber by outside air. Three blower fans (76 cm x 76 cm) are located at the basal part of the OTC and work as CO₂ mixers. The CO₂ gas was injected from nozzles mounted on the inner wall and blower fans then circulated the CO₂ inside the OTC at low speed (Figure 3). The blower fans distribute the high CO₂ concentration and maintain inside air temperature evenly throughout the oil palm canopy. Each OTC is equipped with a gas regulator and a flow meter.

CARBON DIOXIDE CONTROL SYSTEM

Liquid CO₂ was transferred through a vaporiser, a regulator and finally distributed inside the OTC (Figure 4). Vaporised CO₂ was distributed evenly

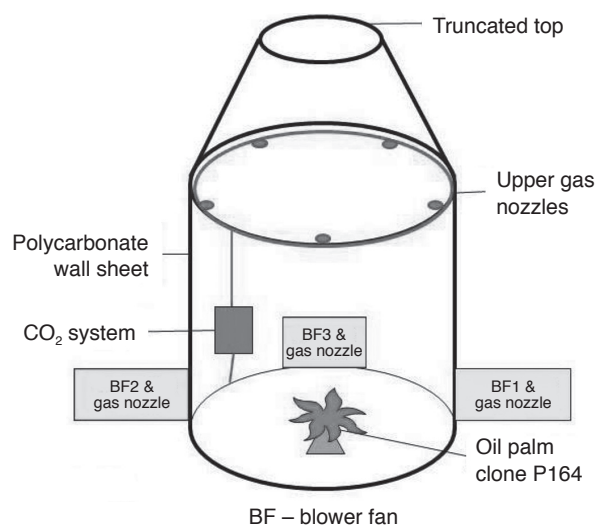


Figure 3. Open Top Chamber diagram with three blower fans, a truncated top and carbon dioxide system.

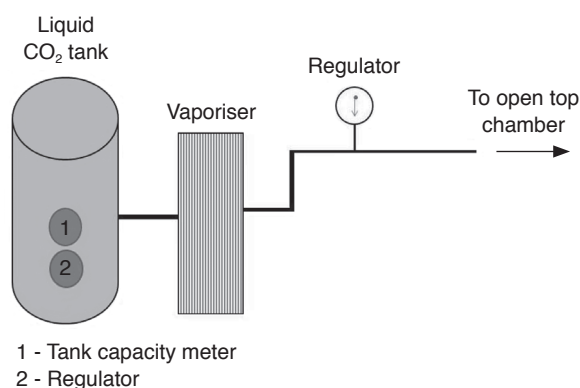


Figure 4. Liquid carbon dioxide goes through a vaporiser and a regulator before being distributed to four units of Open Top Chambers.

via four channels in each chamber *i.e.*, Blower Fan 1, Blower Fan 2, Blower Fan 3 and upper gas nozzles through flow meters (Figure 5). The CO₂ at 99.8% purity was injected through a pressure regulator into the chambers at 10 litres min⁻¹ during active photosynthesis period (8.00 to 11.00 am). During enrichment, three units of blower fan ran at low speed (<0.5 m s⁻¹) to disperse CO₂ evenly throughout the chamber. This is to ensure that the leaf boundary layer is removed and to increase CO₂ diffusion into the leaf mesophyll (Aldrich and Bartok, 1994).

ASSESSMENT OF ENVIRONMENTAL CONDITIONS WITHIN OPEN TOP CHAMBER

The CO₂ level within all OTC was monitored by a CO₂ gas analyser (LI-820, LICOR, USA) which directly analyse air sampled by a micropump unit. Photosynthetically active radiation (PAR) was measured using a quantum sensor (LI-190, LICOR, USA). This data is important to determine

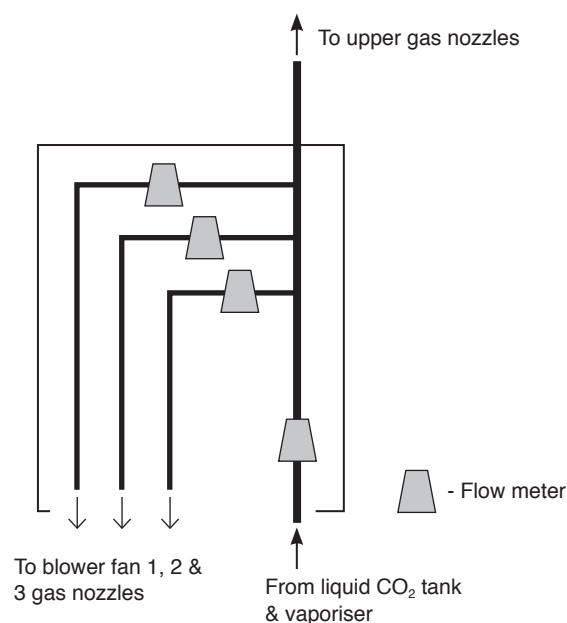


Figure 5. Vaporised carbon dioxide gas flow to upper gas nozzles and three units of blower fan gas nozzles in each Open Top Chamber.

the difference in incoming light within OTC and outside ambient light condition (Messerli *et al.*, 2015). Air temperature, relative humidity and PAR were continuously measured at oil palm canopy height (Campbell Scientific, 1982) in each OTC. The sensor height can be adjusted manually as the palm grows taller and to avoid shading effects from the OTC concrete structure. All installed sensors (CO₂, temperature, relative humidity and PAR) were mounted on a pole with an adjustable arm. Data were recorded at 1 hr intervals and can be retrieved from the control system, *i.e.* Supervisory Control and Data Acquisition (SCADA).

Results show that the OTC was able to maintain a higher CO₂ level inside the OTC during daytime (Figure 6a). The CO₂ concentration was measured at a central location near the oil palm canopy. Air temperature inside the OTC was slightly higher up to 5% (Figure 6b), relative humidity was slightly lower and PAR diminished by as much as 40% (Figures 6c and 6d). It is clear that OTC alters micro-environment compared to unchambered plots particularly temperature, light intensity, relative humidity and wind velocity (Paul and Bert, 1993).

CONCLUSION

A comparison of parameters measured inside the OTC and ambient provides a measure of the chamber effects. However, the effects could have either positive or negative effects on plant growth. For instance, shading effect by the OTC wall could limit light intensity. The increase in air temperature inside the OTC could cause plant water deficits.

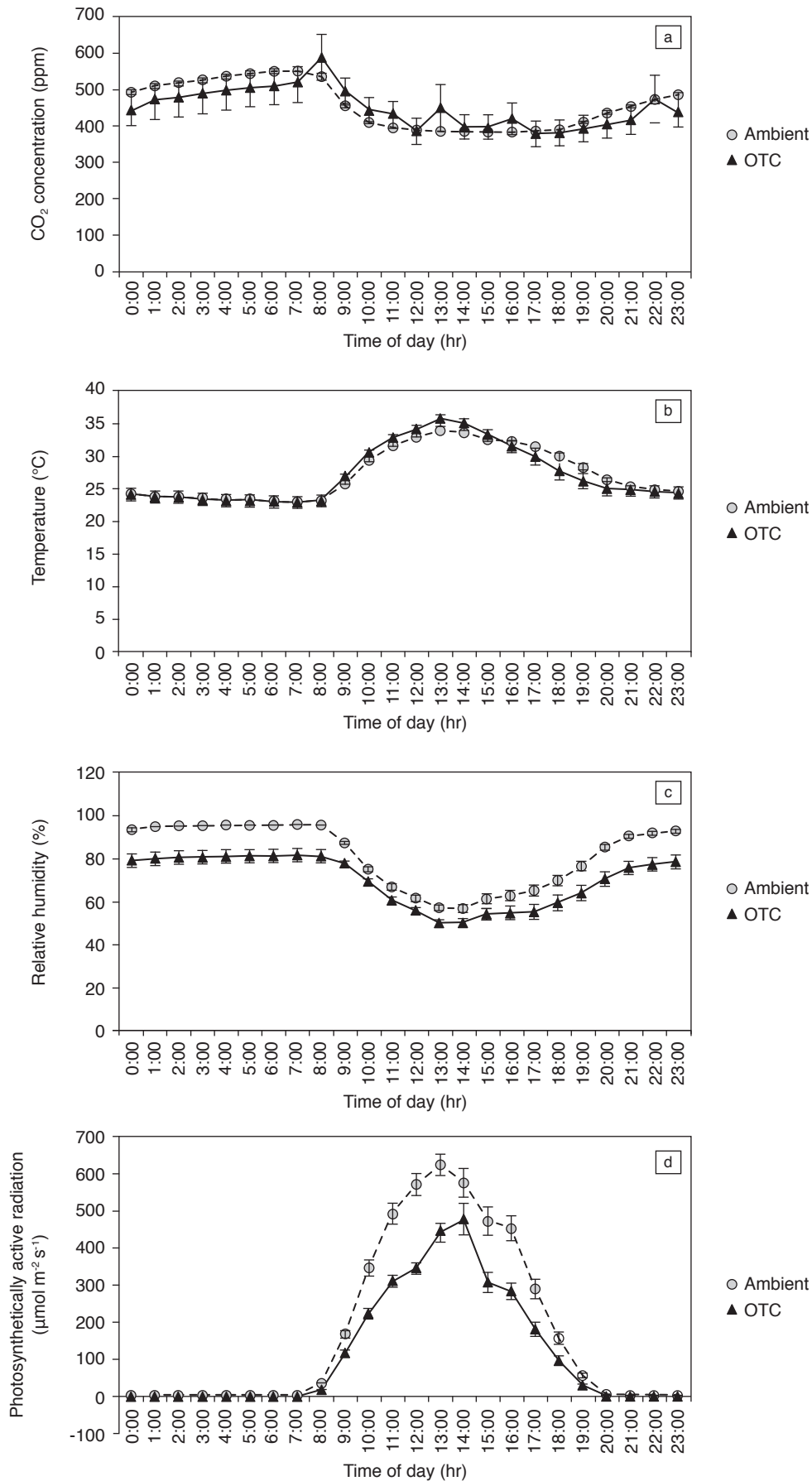


Figure 6. Diurnal pattern of (a) carbon dioxide concentration, (b) air temperature, (c) relative humidity and (d) photosynthetically active radiation inside the Open Top Chamber (OTC) and ambient. Each point represents an average of four weeks data on daily basis (n = 744). Error bar represents standard error of mean.

The OTC method has some limitations and is only appropriate for studying individual plant physiological responses to elevated CO₂. Further investigation is required to explore its suitability for oil palm.

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REFERENCES

- Adams, R M; Rosenzweig, C; Peart, R M; Ritchie, J T; Mccarl, B A; Glycer, J D; Curry, R B; Jones, J W; Boote, K J and Allen, L H Jr (1990). Global climate change and US agriculture. *Nature*, 345: 219-224.
- Aldrich, R A and Bartok, J W (1994). Greenhouse engineering. *Carbon Dioxide Enrichment* (Sailus, M; Chris, N and Sanders, M eds.). University of Connecticut Publication, Ithaca, New York.
- Allen, L H Jr and Prasad, P V V (2004). Crop responses to elevated carbon dioxide. *Encyclopaedia of Plant and Crop Science*. Marcel Dekker Inc., New York. p. 346-348.
- Baker, J T and Allen, L H Jr (1994). Assessment on the impact of rising carbon dioxide and other potential climate changes on vegetation. *Environmental Pollution*, 83: 223-235.
- Barnett, T P; Adam, J C and Lettenmaier, D P (2005). Potential impacts of warming climate on water availability in snow-dominated regions. *Nature*, 438: 303-309.
- Campbell Scientific (1982). *Campbell Scientific Instruction Manual for LI190SB Quantum Sensor Revision 2/15*. Campbell Scientific Incorporated, Utah, USA.
- Drake, B G; Leadley, P W; Arp, W J; Nassiry, D and Curtis, P S (1989). An open top chamber for field studies of elevated atmospheric CO₂ concentration on salt marsh vegetation. *Functional Ecology*, 3(2): 363-371.
- Environmental Protection Agency of the United States (EPA) (2017). Greenhouse gas emissions - Overview of greenhouse gases. <http://www.epa.gov.my/ghgemissions/overview-greenhouse-gases#carbon-dioxide>, accessed on 29 March 2017.
- Ibrahim, M H; Jaafar, H Z E; Haniff, M H and Yusof, M R (2010). Changes in growth and photosynthesis pattern of oil palm (*Elaeis guineensis* Jacq.) seedlings exposed to short-term CO₂ enrichment in a closed chamber. *Acta Physiologiae Plantarum*, 32: 305-313.
- Intergovernmental Panel on Climate Change (IPCC) (2007). The physical science basis. *Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, UK. 976 pp.
- Machova, K (2010). Open top chamber and free air CO₂ enrichment - Approaches to investigate tree responses to elevated CO₂. *iForest – Biogeosciences and Forestry*, 3: 102-105.
- Marschner, H (1995). *Mineral Nutrition of Higher Plants*. Second edition. Academic Press, London, United Kingdom. 889 pp.
- Mccracken, M C (2008). Prospects for future climate change and the reasons for early action. *J. Air and Waste Management Association*, 58: 735-786.
- Messerli, J; Bertrand, A; Bourassa, J; Bèlanger, G; Castonguay, Y; Tremblay, G; Baron, V and Seguin, P (2015). Performance of low-cost open-top chambers to study long-term effects of carbon dioxide and climate under field conditions. *Agronomy J.*, 107(3): 916-920.
- Paul, W L and Bert, G D (1993). Open top chambers for exposing plant canopies to elevated CO₂ concentration for measuring net gas exchange. *Vegetatio*, 104(3): 3-15.
- Pidwirny, M (2006). The carbon cycle. *Fundamental of Physical Geography*. Second edition. University of British Columbia, Canada. 372 pp.
- Ziska, L H (2008). Rising atmospheric carbon dioxide and plant biology: The overlooked paradigm. *Controversies in Science and Technology, From Climate to Chromosomes* (Kleinman, D L and Cloud-Hansen, K A eds.). New Rochele: Liebert Incorporated. p. 379-400.