

GAS EXCHANGE CHARACTERISTICS AND CHLOROPHYLL PIGMENT OF OIL PALM SEEDLINGS UNDER INFLUENCE OF BIOINOCULANTS

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ABSTRACT

The performance of bioinoculants and chemical fertilisers on net photosynthetic rate (P_n), transpiration rate (E), stomatal conductance (G_s), inter cellular CO_2 concentration (C_i), chlorophyll, reducing sugars and total phenols in leaves and total seedling dry matter was assessed in oil palm seedlings grown in nursery for 12 months. Bioinoculants viz. *Azotobacter chroococcum*, *Azospirillum brasilense*, *Bacillus megaterium*, *Frateuria aurantia*, and *Glomus aggregatum* were used individually, combinably and integrated with chemical fertilisers. There were significant differences among the treatments used in experiment for various physiological and biochemical characters. Of all the treatments, integrated use of microbial fertilisers with 25% of recommended dose of chemical fertilisers (RDF) has emerged as the best promising treatment in influencing the above mentioned characters. The results of the study clearly indicated that minimal dose of chemical fertilisers (25% RDF) may be required for exploiting the best possible growth benefits from bioinoculants.

Keywords: oil palm seedlings, bioinoculants, photosynthesis, chlorophyll, reducing sugars, phenols.

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INTRODUCTION

Oil palm (*Elaeis guineensis* Jacq.) is a perennial crop with economic life span of 30 years. The growth and yield of oil palm mainly depend on the quality of seedlings produced in the nursery. Being a heavy feeder, nutrition at the nursery stage is of paramount importance for establishment of healthy and productive commercial oil palm plantations. Microbial inoculants are promising components for integrated solutions to agro-environmental problems because bioinoculants possess the capacity to promote plant growth, enhance nutrient availability and uptake and support the plant health

by improving the microflora (Han and Lee, 2005). Hence, there is a great need for suitable substitute for inorganic fertilisers which are seriously upsetting soil fertility and soil health.

Azospirillum inoculation increased photosynthetic rate of oil palm seedlings as compared to that of control (Amir *et al.*, 2001). Aseri and Rao (2005b) reported that efficient strains of nitrogen fixing bacteria (*Azotobacter chroococcum* and *Azospirillum brasilense*) and AMF-*Glomus* sp. were found to improve the chlorophyll content in leaves of pomegranate seedlings during nursery stage. Rhizobacteria (*Azospirillum* and *Bacillus* sp.) inoculated oil palm seedlings showed positive response in enhancing root and top dry matter under field nursery conditions (Amir *et al.*, 2005). Gas exchange measurements and biochemical parameters are highly useful for comparing and understanding growth and vigour of oil palm seedlings. As no efforts in this line of work in India, the present study was conducted to evaluate influence of microbes on physiological and biochemical parameters of oil palm seedlings.

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MATERIALS AND METHODS

The present investigation was carried out at the Indian Institute of Oil Palm Research, Pedavegi, Andhra Pradesh State, India for two consecutive years. The experimental site is located at 16° 43'N and 81° 09'E with a mean sea level of 13.41 m. Average annual temperature ranged from 21.8°C to 34.8°C in the first year and from 19.6°C to 36.7°C during the second year. Average relative humidity was 69.3% during the first year and 67.7% in the second year. Total amount of rainfall received was 1813.7 mm and 1026 mm during the first and second years, respectively.

Experimental Details

The nursery trial was laid out in completely randomised design (CRD) with 11 treatments replicated five times and 30 seedlings in each treatment. Treatments used were T₁-*Azotobacter chroococcum*, T₂-*Azospirillum brasilense*, T₃-*Bacillus megaterium*, T₄-*Frateuria aurantia*, T₅-*Glomus aggregatum*, T₆-Consortium of bioinoculants, T₇-Consortium of bioinoculants+25% recommended dose of chemical fertilisers (RDF), T₈-Consortium of bioinoculants +50% RDF, T₉-Consortium of bioinoculants+75% RDF, T₁₀-100% RDF and T₁₁-Control without bioinoculants and chemical fertilisers. Commercially, 30 g nitrogen (N), 38 g phosphorus (P) and 25 g potassium (K)/seedling/year are applied to oil palm nursery in India. Tank silt and cattle manure mixed in 2:1 ratio (v/v) was used as a potting mixture (Table 1) for raising oil palm nursery for 12 months in double stage nursery system *i.e.*, primary stage for four months under ultraviolet (UV) stabilised high density polyethylene (HDPE) agro shade net house with 50% shade and secondary stage for eight months in open condition.

TABLE 1. PHYSICO-CHEMICAL CHARACTERISTICS OF POTTING SOIL USED FOR RAISING OIL PALM NURSERY

Sand (%)	54.67
Silt (%)	8.35
Clay (%)	36.98
Texture	Clayey
pH	7.12
Electrical conductivity (EC) (dS m ⁻¹)	0.46
Organic carbon (OC) (%)	1.25
Phosphorus (P) (ppm)	24.62
Potassium (K) (ppm)	145.65
Concentration (Ca) (100 g meq ⁻¹)	3.34
Magnesium (Mg) (100 g meq ⁻¹)	2.16

Oil palm seedlings were raised in polybags of 23 cm x 15 cm during the primary stage and 45

cm x 38 cm in secondary stage. Uniform, healthy and 65 days old oil palm seed sprouts of *Tenera* hybrid combination (1140 *Dura* x 1988 *Pisifera*) were used as planting material during both the years. Bioinoculants were applied thrice at four months interval whereas chemical fertilisers Di-ammonium phosphate (DAP) and complex (17:17:17) were applied manually to the nursery at monthly interval. Nursery operations like watering and weeding were carried out uniformly for all the treatments. There were no serious insect pests and diseases during the nursery period.

Lignite-based *Azotobacter chroococcum* (1x10⁸ cfu g⁻¹), *Azospirillum brasilense* (1x10⁸ cfu g⁻¹), *Bacillus megaterium* (1x10⁸ cfu g⁻¹), *Frateuria aurantia* (1x10⁸ cfu g⁻¹) and soil-based *Glomus aggregatum* (800 infectious propagules g⁻¹) were applied to potting mixture at the time of planting of seed sprouts, shifting of primary seedlings to secondary stage bags and finally eight months after planting of sprouts and the quantity of microbial inoculants per bag used was 10 g, 25 g and 5 g, respectively. In the combined treatment, *Bacillus megaterium*, *Frateuria aurantia* and *Glomus aggregatum* with above mentioned doses while *Azotobacter chroococcum* (5, 12.5 and 2.5 g) and *Azospirillum brasilense* (5, 12.5 and 2.5 g) with 50% of the above mentioned doses per bag were mixed together and applied to the potting mixture.

Physiological Observations

Dry matter of oil palm seedlings was recorded at four stages with three months interval *i.e.*, stage-1 after three months, stage-2 after six months, stage-3 after nine months and stage-4 after 12 months at the nursery. Seedlings were separated into root, stem and leaf and recorded as fresh weight of roots (g), stem (g) and leaf (g) portions at four stages. Then samples were kept in hot air oven at 60°C and dry weight of plant parts and total dry matter was estimated by using top pan electronic balance. Gas-exchange measurements like net photosynthetic rate (P_n), transpiration (E), stomatal conductance (G_s) and inter cellular CO₂ concentration (C_i) were recorded at stage-2 and stage-4 by using a Portable Photosynthesis System (LI-COR Biosciences, Lincoln, USA) connected to a PLC-4 (6.25 cm²) leaf chamber. During measurements, incident photon flux density was 800 μmol (photon) m⁻² s⁻¹, leaf temperature 25°C and ambient CO₂ concentration (C_a) 360 μmol mol⁻¹. Measurements were made on fully opened, matured and healthy leaf (third leaf from top of the canopy) under bright sunlight between 9.00-11.00 am.

Biochemical Analysis

Biochemical compounds like chlorophyll (chlorophyll *a*, *b* and total chlorophyll), reducing

sugars and total phenols in leaves were quantified at stage-2 (six months) and stage-4 (12 months) of oil palm nursery by using methods of Hiscox and Israelstam (1979), Somogyi (1952) and Malik and Singh (1980), respectively.

Statistical Analysis

Treatment effects were assessed for individual years using one way analysis of variance (ANOVA) and pooled effects were assessed using repeated measures ANOVA. Significance levels were tested at $p \leq 0.5$. Post-hoc analysis was done using Least Significant Difference (LSD) test. All the statistical analysis was carried out using generalised linear model (GLM) procedure statistical software SAS version 9.3.

RESULTS AND DISCUSSION

Gas Exchange Parameters

There were significant differences among the treatments and all the treatments were found significantly superior to the control for Pn, E, Gs and Ci in leaves of oil palm seedlings (Table 2). Of all the treatments, T₇-Consortium of bioinoculants+25% RDF recorded the highest Pn (13.85 $\mu\text{mol m}^{-2} \text{s}^{-1}$)

which was highly significant to other treatments except T₈ (13.24 $\mu\text{mol m}^{-2} \text{s}^{-1}$), T₅ (12.51 $\mu\text{mol m}^{-2} \text{s}^{-1}$) and T₆ (12.49 $\mu\text{mol m}^{-2} \text{s}^{-1}$). Similar results were noticed in egg plant (Han and Lee, 2005) and banana plantlets grown under hydroponic condition (Baset Mia *et al.*, 2009). The higher Pn at light saturation was related to the higher efficiency of ribulose biphosphate (RuBp) regeneration capacity of the host plant. This phenomenon is linked to higher Gs, higher inter cellular CO₂ concentration and maximal interception of light through increased leaf surface and minimal carbon loss in the dark respiration process (Henson, 1991). Bondada and Oosterius (1998) have pointed out that Pn of the host plant is related to N content of leaf. The higher N in inoculated seedlings also might have contributed to the formation of chlorophyll which consequently might have increased the photosynthetic activity. Increased Pn (Mathur and Vyas, 1995) can be attributed to enhanced leaf area and chlorophyll content and vigorous growth of ber seedlings.

Among individual microbial fertilisers, maximum Pn was achieved in T₅-*Glomus aggregatum* (12.51 $\mu\text{mol m}^{-2} \text{s}^{-1}$) which was relatively better than T₆-Consortium of bioinoculants (12.49 $\mu\text{mol m}^{-2} \text{s}^{-1}$) and other individual bioinoculants. Enhanced photosynthetic rate in T₅-*Glomus aggregatum* may be due to higher chlorophyll and carotenoid content in leaves (Mathur and Vyas, 1999; Krishna

TABLE 2. EFFICACY OF BIOINOCULANTS AND CHEMICAL FERTILISERS ON GAS EXCHANGE PARAMETERS OF OIL PALM SEEDLINGS GROWN IN NURSERY

Treatment	Photosynthetic rate ($\mu\text{mol m}^{-2} \text{s}^{-1}$)			Transpiration rate ($\text{mmol m}^{-2} \text{s}^{-1}$)			Stomatal conductance ($\text{mmol m}^{-2} \text{s}^{-1}$)			Inter cellular CO ₂ concentration (ppm)		
	1 yr	2 yr	Pooled mean	1 yr	2 yr	Pooled mean	1 yr	2 yr	Pooled mean	1 yr	2 yr	Pooled mean
T ₁	11.62	12.84	12.23	6.09	6.09	6.09	0.20	0.17	0.18	250.79	218.75	234.77
T ₂	11.77	12.18	11.97	6.50	5.89	6.20	0.22	0.17	0.19	250.43	208.97	229.70
T ₃	11.18	11.67	11.42	6.58	6.30	6.44	0.23	0.18	0.20	247.66	244.25	245.95
T ₄	11.45	11.17	11.31	6.56	6.34	6.45	0.20	0.19	0.19	248.88	201.77	225.32
T ₅	12.24	12.78	12.51	6.95	7.56	7.26	0.22	0.23	0.22	257.62	247.88	252.75
T ₆	12.01	12.97	12.49	8.34	7.05	7.69	0.26	0.20	0.23	271.26	233.88	252.57
T ₇	13.37	14.35	13.85	8.55	8.00	8.27	0.27	0.21	0.24	264.83	260.32	262.57
T ₈	12.96	13.53	13.24	8.29	7.72	8.00	0.24	0.22	0.22	261.12	249.84	255.48
T ₉	12.34	11.42	11.88	7.57	7.04	7.30	0.20	0.14	0.17	255.02	229.68	242.35
T ₁₀	11.72	11.99	11.85	6.54	6.75	6.64	0.18	0.13	0.15	237.76	227.21	232.48
T ₁₁	10.24	9.09	9.66	5.65	5.92	5.78	0.18	0.14	0.15	221.42	207.94	214.68
LSD-5%	1.07	1.65	1.38	0.77	0.58	0.92	0.03	0.02	0.03	15.02	26.92	17.90
SEM	0.92	1.42	1.70	0.66	0.49	1.14	0.02	0.02	0.04	12.89	23.10	22.13

Note: T₁-*Azotobacter chroococcum*, T₂-*Azospirillum brasilense*, T₃-*Bacillus megaterium*, T₄-*Frateuria aurantia*, T₅-*Glomus aggregatum*, T₆-Consortium of bioinoculants, T₇-Consortium of bioinoculants+25% RDF, T₈-Consortium of bioinoculants+50% RDF, T₉-Consortium of bioinoculants+75% RDF, T₁₀-100% RDF and T₁₁-Control. RDF - recommended dose of chemical fertilisers, LSD - Least Significant Difference, SEM - scanning electronic microscopy.

et al., 2006). In addition, arbuscular mycorrhizal fungi (AMF) may function as a metabolic sink causing basipetal mobilisation of photosynthates to roots thus providing a stimulus for greater photosynthetic activity (Bevege *et al.*, 1975). Nowak (2004) and Wu and Xia (2006) reported increased rate of photosynthesis with mycorrhiza association in geranium and citrus seedlings, respectively as compared with the control.

Treatment T₇-Consortium of bioinoculants+25% RDF (8.27 mmol m⁻² s⁻¹) had shown the highest E which was significantly superior to other treatments except T₈-Consortium of bioinoculants+50% RDF (8.00 mmol m⁻² s⁻¹) and T₆-Consortium of bioinoculants (7.69 mmol m⁻² s⁻¹). Among individual bioinoculants, only the treatment T₅-*Glomus aggregatum* (7.26 mmol m⁻² s⁻¹) differed significantly with the control (5.78 mmol m⁻² s⁻¹). The present results are in agreement with the findings of Yano Melo *et al.* (1999) in banana plantlets. Increase in E might be from enhanced metabolic activity associated with AMF colonisation (Johnson and Hammel, 1985), better uptake of water as mycorrhiza in rhizosphere explores the soil better than un-inoculated roots. Further, improved water transport under well irrigated conditions may be correlated with increased P nutrition provided by AMF (Graham and Syversten, 1985).

Among the treatments, T₇-Consortium of bioinoculants+25% RDF (0.24 mmol m⁻² s⁻¹) recorded the maximum G_s and it was markedly higher than other treatments except T₆ (0.23 mmol m⁻² s⁻¹), T₈ (0.22 mmol m⁻² s⁻¹) and T₅ (0.22 mmol m⁻² s⁻¹). There were no significant differences among T₅-*Glomus aggregatum* (0.22 mmol m⁻² s⁻¹), T₆-Consortium of bioinoculants (0.23 mmol m⁻² s⁻¹), T₇-Consortium of bioinoculants+25% RDF (0.24 mmol m⁻² s⁻¹) and T₈-Consortium of bioinoculants+50% RDF (0.22 mmol m⁻² s⁻¹) for G_s in leaves. Increased G_s in inoculated seedlings may be ascribed to higher N content in leaf (Bondada and Oosterius, 1998). Similarly, Nowak (2004) observed increased G_s in geranium under mycorrhisation.

Highest inter cellular CO₂ concentration value in leaves was noticed in T₇-Consortium of bioinoculants+25% RDF (262.57 ppm) which was markedly superior to T₁₁-Control (214.68 ppm) and T₁₀-100% RDF (232.48 ppm). Among individual bioinoculants, the best results for inter cellular CO₂ concentration in leaves were noted with T₅-*Glomus aggregatum* (252.75 ppm) followed by T₃-*Bacillus megaterium* (245.95 ppm). The effect of T₅-*Glomus aggregatum* (252.75 ppm) can be compared with T₆-Consortium of bioinoculants (252.57 ppm) and integrated use of bioinoculants+chemical fertilisers. Further, the treatment T₅-*Glomus aggregatum* (252.75 ppm) exhibited significant performance over T₁₀-100% RDF (232.48 ppm).

Dry Matter Production

The treatment T₇-Consortium of bioinoculants+25% RDF and T₁₁-Control recorded the maximum (200.53 g) and minimum (102.91 g) dry matter among others (Table 3). Significant enhancement in seedling dry matter under T₇-Consortium of bioinoculants+25% RDF might be due to vigorous growth of seedlings with more number of leaves and leaflets and root density which is certainly due to greater absorption of nutrients, higher P_n, metabolic activities (Mathur and Vyas, 1999) and accumulation of nutrients in plant tissues. Findings of the present study are confirmed with reports of Aseri and Rao (2005a) in ber seedlings and Noor Aishah *et al.* (2009) in oil palm seedlings.

Treatment T₁₀-100% RDF had shown comparatively better results over individual bioinoculants and it was found on par with T₆-Consortium of bioinoculants and T₈-Consortium of bioinoculants+50% RDF and T₉-Consortium of bioinoculants+75% RDF. Similarly, Azlin *et al.* (2009) noticed the superiority of chemical fertilisers over individual application of bioinoculants in tissue cultured oil palm plants. Increased dry matter production can be strongly correlated with improved accumulation of N due to NFB, P due to phosphate solubilising bacteria (PSB) and AMF and K due to potash mobilise (Ratha Krishnan *et al.*, 2004) in *Simarouba glauca* seedlings. The same trend was reported in *Solanum viarum* nursery (Hemashenpagam and Selvaraj, 2011) and pomegranate cuttings (Muzaffar Mir *et al.*, 2012). Among individual treatments, T₅-*Glomus aggregatum* was found on par with T₁₀-100% RDF and T₆-Consortium of bioinoculants for dry matter production. Similarly, Nagaveni *et al.* (1998) in *Tectona grandis* reported that the growth of AMF inoculated seedlings can be compared with inorganic fertilisers.

Chlorophyll

Treatment T₇-Consortium of bioinoculants+25% RDF excelled among others by recording the maximum quantity of chlorophyll *a* (1.13 mg g⁻¹), chlorophyll *b* (0.50 mg g⁻¹) and total chlorophyll (1.63 mg g⁻¹) in leaves while the minimum amount was estimated under the control (Table 3). Among individual treatments, higher values of chlorophyll *a* (0.85 mg g⁻¹), chlorophyll *b* (0.44 mg g⁻¹) and total chlorophyll content (1.29 mg g⁻¹) in leaves were manifested in T₅-*Glomus aggregatum* which was significantly superior to the control and the rest of individual treatments and it was found on par with T₆-Consortium of bioinoculants and T₁₀-100% RDF. Similar results were obtained in pomegranate

TABLE 3. EFFICACY OF BIOINOCULANTS AND CHEMICAL FERTILISERS ON DRY MATTER PRODUCTION AND CHLOROPHYLL CONTENT OF OIL PALM SEEDLINGS GROWN IN NURSERY

Treatment	Total dry matter (g)			Chlorophyll a (mg g ⁻¹)			Chlorophyll b (mg g ⁻¹)			Total chlorophyll (mg g ⁻¹)		
	1 yr	2 yr	Pooled mean	1 yr	2 yr	Pooled mean	1 yr	2 yr	Pooled mean	1 yr	2 yr	Pooled mean
T ₁	161.54	118.28	139.91	0.73	0.73	0.73	0.31	0.38	0.34	1.03	1.10	1.07
T ₂	147.72	120.08	133.90	0.75	0.70	0.72	0.30	0.49	0.39	1.05	1.19	1.12
T ₃	156.38	126.04	141.21	0.79	0.78	0.78	0.27	0.43	0.35	1.05	1.22	1.13
T ₄	145.75	108.04	126.90	0.79	0.78	0.78	0.27	0.34	0.30	1.06	1.11	1.08
T ₅	166.88	140.94	153.91	0.89	0.81	0.85	0.38	0.52	0.44	1.26	1.32	1.29
T ₆	170.82	156.58	163.70	1.03	0.87	0.95	0.35	0.58	0.46	1.38	1.45	1.41
T ₇	232.52	168.54	200.53	1.17	1.10	1.13	0.46	0.54	0.50	1.63	1.64	1.63
T ₈	202.37	149.86	179.37	1.17	0.89	1.03	0.40	0.48	0.44	1.57	1.37	1.47
T ₉	201.07	143.36	172.21	1.01	0.82	0.91	0.36	0.44	0.40	1.36	1.25	1.31
T ₁₀	188.98	136.75	162.86	0.84	0.77	0.80	0.34	0.39	0.36	1.18	1.16	1.16
T ₁₁	137.51	68.31	102.91	0.69	0.71	0.70	0.30	0.29	0.29	0.99	1.00	0.99
LSD-5%	13.65	23.52	23.43	0.09	0.10	0.13	0.04	0.05	0.07	0.10	0.11	0.15
SEM	16.82	28.98	41.21	0.08	0.09	0.16	0.03	0.04	0.09	0.08	0.09	0.18

Note: T₁-*Azotobacter chroococcum*, T₂-*Azospirillum brasilense*, T₃-*Bacillus megaterium*, T₄-*Frateuria aurantia*, T₅-*Glomus aggregatum*, T₆-Consortium of bioinoculants, T₇-Consortium of bioinoculants+25% RDF, T₈-Consortium of bioinoculants+50% RDF, T₉-Consortium of bioinoculants+75% RDF, T₁₀-100% RDF and T₁₁-Control.

RDF - recommended dose of chemical fertilisers, LSD - Least Significant Difference, SEM - scanning electronic microscopy.

nursery (Aseri and Rao, 2005b), guava seedlings (Panneerselvam *et al.*, 2012) and *Marsdenia volubilis* seedlings (Sandhya *et al.*, 2013). Higher stomatal conductance, photosynthesis and transpiration must be responsible for enhanced chlorophyll in AMF inoculated plants (Krishna and Bagyaraj, 1984).

Overall, chlorophyll *a*, chlorophyll *b* and total chlorophyll contents in oil palm leaves were the maximum under integrated use of bioinoculants+chemical fertilisers. The present study confirms the reports of Aseri and Rao (2005a) in ber seedlings and Noor Aishah (2009) in oil palm seedlings. Improved chlorophyll content in inoculated seedlings is an indication of enhanced Mg, Fe and Cu uptake which are essential for biosynthesis of chlorophyll in plants. Azlin *et al.* (2009) reported that plant-bacteria association enhances chlorophyll of tissue cultured oil palm plantlets due to indole acetic acid (IAA) production. Cytokinin produced by microbial inoculants might have become a sink to attract nutrients like Mg, Fe and K which in turn might have resulted in greater synthesis of chlorophyll (Lalitha *et al.*, 2004) in betel vine. An increase in chlorophyll content was ascribed to enhanced availability of water and minerals (Shaban and Mohsen, 2009). Significantly higher total chlorophyll content in leaves of inoculated seedlings might have resulted from enhanced stomatal conductance, carbon assimilation (Levy and Krikun, 1980) and transpiration (Hayman,

1983), plant growth and biomass production (Kolher *et al.*, 2007; Panneerselvam *et al.*, 2012).

Reducing Sugars

Maximum quantity of reducing sugars (Table 4) was estimated under integrated application of bioinoculants particularly with T₇-Consortium of bioinoculants+25% RDF (5.08 mg g⁻¹) and then followed by T₅-*Glomus aggregatum* (4.65 mg g⁻¹), T₆-Consortium of bioinoculants (3.52 mg g⁻¹) and T₁₀-100% RDF (3.30 mg g⁻¹) when compared with the control (2.45 mg g⁻¹). Among individual bioinoculants, T₅-*Glomus aggregatum* (4.65 mg g⁻¹) recorded the highest quantity of reducing sugars in leaves and it was also significantly superior to other treatments except T₇-Consortium of bioinoculants+25% RDF (5.08 mg g⁻¹). This observation is corroborated with Aseri *et al.* (2008) who reported higher level of reducing sugars in leaves of pomegranate seedlings. Increased plant growth and biomass production and higher Pn might have boosted the production of reducing sugars in leaves of inoculated *Lactuca sativa* seedlings (Kolher *et al.*, 2007).

Total Phenols

Of all the treatments, maximum and minimum quantity of total phenols (Table 4) in leaves was

TABLE 4. EFFICACY OF BIOINOCULANTS AND CHEMICAL FERTILISERS ON REDUCING SUGARS AND TOTAL PHENOLS CONTENT IN LEAVES OF OIL PALM SEEDLINGS GROWN IN NURSERY

Treatment	Reducing sugars (mg g ⁻¹)			Total phenols (µg g ⁻¹)		
	1 yr	2 yr	Pooled mean	1 yr	2 yr	Pooled mean
T ₁	3.28	3.10	3.19	3.80	1.32	2.56
T ₂	3.87	3.07	3.47	3.30	1.29	2.29
T ₃	3.39	3.05	3.22	2.93	1.56	2.24
T ₄	2.81	2.64	2.72	3.08	1.30	2.19
T ₅	5.27	4.03	4.65	3.15	2.06	2.60
T ₆	3.70	3.34	3.52	3.43	2.18	2.81
T ₇	5.58	4.58	5.08	4.37	2.77	3.57
T ₈	3.58	3.79	3.68	4.05	2.45	3.25
T ₉	3.50	3.41	3.45	3.45	2.13	2.79
T ₁₀	3.51	3.10	3.30	3.96	1.86	2.91
T ₁₁	2.68	2.21	2.45	3.47	1.33	2.40
LSD-5%	0.25	0.26	0.31	0.21	0.14	0.22
SEM	0.21	0.22	0.37	0.18	0.12	0.26

Note: T₁-*Azotobacter chroococcum*, T₂-*Azospirillum brasilense*, T₃-*Bacillus megaterium*, T₄-*Frateuria aurantia*, T₅-*Glomus aggregatum*, T₆-Consortium of bioinoculants, T₇-Consortium of bioinoculants+25% RDF, T₈-Consortium of bioinoculants+50% RDF, T₉-Consortium of bioinoculants+75% RDF, T₁₀-100% RDF and T₁₁-Control.

RDF - recommended dose of chemical fertilisers, LSD - Least Significant Difference, SEM - scanning electronic microscopy.

recorded with T₇-Consortium of bioinoculants+25% RDF (3.57 µg g⁻¹) and T₄-*Frateuria aurantia* (2.19 µg g⁻¹), respectively. Higher phenol content was quantified in integrated application of bioinoculants+chemical fertilisers which were distinctly superior to the control. Similarly, results under T₆-Consortium of bioinoculants (2.81 µg g⁻¹) and T₁₀-100% RDF (2.91 µg g⁻¹) were significantly better than the control (2.40 µg g⁻¹). The present results find support from the reports of Panneerselvam *et al.* (2012), Ramakrishnaiah and Vijaya (2013) and El-Quesni *et al.* (2013) who noticed significant improvement in phenols production in leaves/shoots of guava, pomegranate and jatropha nursery, respectively under combined use of bioinoculants in comparison with the control. Enhanced phenolic content in plant tissue can be ascribed to increased polyphenol oxidase activity (Krishna *et al.*, 2006) and the use of accumulated nitrate in plant thus enabling it to use more carbohydrates for structural growth under the influence of biofertilisers (Hanafy Ahmed *et al.*, 2000). Further, higher phenol content in inoculated seedlings might have resulted from enhanced plant growth and biomass production (Kolher *et al.*, 2007). There were no significant differences for phenol content in leaves between individual bioinoculants and the control and this indicated non-effectiveness of individual microbial fertilisers.

Further, gradual decline in gas exchange parameters, chlorophyll content and dry matter production was observed when the RDF was increased from 25% to 75% under integration.

CONCLUSION

The results of the study demonstrated that inoculation of oil palm nursery seedlings with microbes *vis-à-vis* 100% RDF and the control induced positive changes in gas exchange parameters, chlorophyll pigment, reducing sugars and total phenols which in turn influenced significantly the dry matter of seedlings. The treatment T₅-*Glomus aggregatum* had shown significantly better performance for all the characters studied and it was equally effective when compared with consortium of bioinoculants and 100% RDF. Overall, results clearly illustrated that integrated application of consortium of bioinoculants in combination with 25% RDF can be a recommendation for better growth and performance of oil palm seedlings. Further, this paves a way for reduction of 75% of RDF and make oil palm nursery self-sustainable.

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