INTRODUCTION

Breast cancer (BC) is ranked as the most frequent cancer among women worldwide; with an estimated 11.6% new cases as reported in the global cancer statistics 2018 (Bray et al., 2018). According to the Malaysian National Cancer Registry (2012-2016) report (Azizah et al., 2019), one in 20 women may develop BC in their lifetime and the incidence of BC was highest in Chinese, followed by Indians and Malays (Azizah et al., 2019). The mean age standardised rate (ASR) amongst Malaysian women who developed BC was reported to be 39.3 per 100,000 populations in 2006 (Azizah et al., 2019).

The survival rate of patients with BC is considered crucial as it can control the mortality rate; and in Malaysia, the 5-year survival rate was found to be favourably improved (Abdullah et al., 2013). Use of alternative medicine, particularly dietary supplements were reported to be common practices among Malaysian BC survivors (Shaharudin et al., 2011).

Bioactive compounds derived from natural sources are preferred compared to synthetic medicinal products as these have lesser side-effects. One such compound is tocotrienols (T3),

ABSTRACT

Gamma-tocotrienol (γT3) is an isoform of vitamin E found abundantly in palm oil, which is reported to possess antioxidant and anticancer activities. However, the immune-modulating properties of γT3 have yet to be elucidated. Breast cancer (BC) was induced in female BALB/c mice by injecting 4T1 murine mammary cancer cells into their mammary fat pads. When the tumour was palpable, the animals were randomly assigned into two groups: (i) control [fed twice daily with 50 μL vehicle (soy oil)] or (ii) experimental (fed twice daily with 50 μL of 0.5 mg of γT3). Results show that mice fed with γT3 had reduced tumour growth and metastasis. However, there are no marked changes in the percentages of peripheral blood leukocytes and cytokines production in these animals. Immunohistochemistry using antibodies to murine CD4, IL12Rβ2, IL24 and FoxP3 on tumour sections from γT3 treated mice suggested that γT3 induced suppression of tumour growth and metastasis as well as reduced immunosuppression in the tumour microenvironment.

It can be concluded that, γT3 has the potential to suppress tumour growth and metastasis in this model. Further investigation on the host immune response is possible by prolonging treatment duration against BC.

Keywords: breast cancer, gamma-tocotrienol, mouse, supplementation, 4T1.
a natural isoform of vitamin E, which is found abundantly in palm oil and widely known for its anticancer, antioxidant and other beneficial health effects (Kabir et al., 2017; Montagnani Marelli et al., 2019). Vitamin E is divided into two families, namely tocopherol and T3. Major sources of dietary tocopherols are wheat-germ oil, safflower-seed oil, maize oil, soybean oil, whilst T3 can be found abundantly in palm oil, rice-bran oil and palm kernel oil. Both tocopherols and T3 exist naturally as four analogues i.e., alpha (α), beta (β), delta (δ) and gamma (γ). Vitamin E from palm oil is known as tocotrienol-rich fraction (TRF), which contains all four T3 analogues and alpha-tocopherol (Mba et al., 2015). There is increasing evidence suggesting T3s are potent antioxidants, which can potentially protect against various diseases, including cancer (Abraham et al., 2019; Montagnani Marelli et al., 2019). Evidence from cell-based studies have shown that T3 can inhibit growth and proliferation of various cancer cell lines (Aggarwal et al., 2019; De Silva et al., 2016; Fontana et al., 2020; Ghanem et al., 2019; Nesaretnam et al., 1998). The ability of T3 to selectively inhibit the proliferation of cancer cells strongly support their anticancer potential. Furthermore, T3 is also credited for its immune-modulating activities in a syngeneic mouse model of BC. For instance, TRF supplementation was reported to induce cancer-specific immune responses in a syngeneic mouse model of BC (Hafid et al., 2010). Previous studies had shown that T3 can inhibit both the proliferation of 4T1 mouse mammary cancer cells and tumour growth (Selvduray et al., 2010; Subramaiam et al., 2021).

The focus of this article is to investigate the effects of daily supplementation of gamma-tocotrienol (γT3) to inhibit tumour growth and exert immunomodulation in a syngeneic mouse model of BC. The findings from this preclinical study may be useful to predict and develop new treatment strategies using palm T3 to fight BC.

MATERIALS AND METHODS

Cell Lines and Culture Conditions

The 4T1 mouse mammary cancer cells were obtained from the American Tissue Culture Collection (ATCC) (ATCC, Rockville, USA) and cultured according to the protocol provided by the ATCC. The cells were cultured at 37°C in Roswell Park Memorial Institute Medium (RPMI) medium (GIBCO, Invitrogen, USA) supplemented with L-glutamine (GIBCO, Invitrogen, USA), 10% fetal bovine serum (FBS) (GIBCO, Invitrogen, USA), and 1% penicillin-streptomycin (GIBCO, Invitrogen, USA) in a humidified incubator containing 5% carbon dioxide (CO₂).

Preparation of Treatment

The γT3 used in this study was obtained from the Malaysian Palm Oil Board (MPOB). The γT3 was extracted from palm oil with a purity range of 95%-99% (Maarasyid et al., 2014). The γT3 was prepared fresh every day prior to feeding by dissolving 0.5 mg of γT3 in 50 μL of soy oil, which served as the vehicle.

Experimental Procedure

Female BALB/c mice aged 5-6 weeks (Cheneur Suppliers, Malaysia) were housed at the animal holding facility (AHF), International Medical University (IMU). All experimental procedures were approved by the Joint Committee for Research and Ethics, IMU (IMU-R060-2010) and the study was conducted according to the IMU animal ethics guidelines. The mice were maintained at 25 ± 2°C with 12:12 hr dark/light cycle and they had free access to commercial food pellets and water throughout the study period. After one-week of acclimatisation, six (n=6) mice were sacrificed to collect baseline data. The remaining mice (n=48) received an injection of 100 μL of 4T1 cells (10 000 cells/mL) at their right thoracic mammary fat pad to induce BC (Selvduray et al., 2010). Once tumour was palpable (day 14), the mice were randomly divided into vehicle and treatment groups. The animals in the vehicle group were fed with 50 μL vehicle (soy oil) whilst the mice in the treatment group were fed with 50 μL of vehicle containing 0.5 mg γT3 twice daily (morning and evening) for a period of 35 days by oral gavage. Throughout the study period, six mice (n=6) from each group were sacrificed every seven days (day 14, 21, 28, 35). Tumour size was monitored and measured weekly using a digital calliper. Tumour volume was calculated using the formula: V = 0.52 x L² x W (V: volume, L: length and W: width) (Selvduray et al., 2010). The study was terminated at day 35 when the tumour load in the vehicle group became too big and the animals started to show signs of distress. The animals were humanely euthanised using the cervical dislocation method.

Histopathology Assessment

At autopsy, tumour, lung, kidney and liver were removed and fixed in 10% formalin for 48 hr before it was processed using an automatic tissue processor (Leica TP1020 Automatic Tissue Processor, Leica, Germany). These tissues were paraffin-embedded and sectioned at 4 μm thickness using a rotary microtome (Rotary Microtome, Leica, Germany). The tissue sections were stained with haematoxylin and eosin (H&E) stains (Leica, Germany). The stained slides were evaluated by a pathologist using a light microscope (Nikon eclipse, Japan) to look for extent and signs of metastasis.
**Immunohistochemistry Staining**

Tumours sections taken from vehicle-fed and γT3-fed mice were incubated at room temperature with respective primary antibodies for 1 hr. The primary antibodies used were (i) anti-CD4 [rabbit anti-mouse CD4 polyclonal antibody (Dako, Denmark)]; (ii) anti-Forkhead box P3 (FoxP3) [rabbit anti-mouse FoxP3 polyclonal antibody] (Dako, Denmark); (iii) anti-interleukin-24 (IL-24) [rabbit anti-mouse IL-24 polyclonal antibody] (Dako, Denmark) and (iv) anti-interleukin-12-beta-2 receptor (IL-12Rβ2) [rabbit anti-mouse IL-12Rβ2 polyclonal antibody] (Dako, Denmark). After 1 hr, the slides were washed with a wash buffer and incubated for 20 min at room temperature with a biotinylated secondary antibody [biotinylated anti-rabbit polyclonal antibody] (Dako, Denmark). Following this, the slides were washed and incubated at room temperature for 20 min with streptavidin-conjugated horseradish peroxidase (HRP) (Dako, Denmark). After the slides were washed, a substrate chromogen (Dako, Denmark) was added to the sections and the slides were incubated at room temperature for an additional 20 min. Following this, the slides were counterstained with haematoxylin and viewed under a light microscope. Staining intensity was determined using a semi-quantitative method of Allred scoring system (Fedchenko and Reifenrath, 2014), whereby the percentage of stained cells and intensity of staining were calculated. Each section was given a score based on the staining pattern (0= negative, 1= weak, 2= moderate and 3= strong).

**Leucocyte Subsets**

At autopsy, blood was obtained via cardiac puncture and collected into heparinised tubes. The blood was centrifuged (200 g for 10 min at 4°C) to separate plasma and cells. The plasma was removed, the red blood cells (RBC) were lysed by adding 2 mL of RBC lysis buffer (BD Pharm Lyse, USA) and the leucocytes were recovered by centrifugation (200 g for 5 min at 4°C). Then, the leucocytes were washed with cold phosphate-buffered saline (PBS) and recovered by centrifugation. Following this, the cells were resuspended in a staining buffer (FACS Sheath Fluid BD, USA) and counted. Cell count was adjusted to 1 x 10⁶/ 100 μL cells. The cells were added to appropriately labelled tubes and stained with conjugated antibodies for flow cytometer analysis. The antibodies used were specific to murine (i) TCR-β⁺CD4⁺CD8A⁺ to identify T-lymphocytes (Biosciences BD, USA); (ii) CD335⁺CD3ε⁺CD49b⁺ to detect natural killer (NK) cells (Biosciences BD, USA); and (iii) CD4⁺CD25⁺FR-4⁺ to identify T-regulatory (Treg) cells (Biosciences BD, USA). The cells were stained for 20 min at room temperature in the dark. Following this, the cells were washed with a staining buffer and recovered by centrifugation. The stained cells were resuspended in 0.5 mL of cell fix buffer (Cell Fix BD, USA) and analysed using a multicolour flow cytometer (FACS Calibur, Becton-Dickson, USA). Data was collected using the Cell Quest software provided by the manufacturer (Becton Dickinson, USA). In each acquisition, 10 000 cells were collected for data analysis.

**Quantification of Cytokines**

When the animals were sacrificed, the spleen from each animal was aseptically removed and a splenocyte suspension was prepared. The cells were counted and plated in a 96-well plate at a cell density of 5 x 10⁶ splenocytes/well. The plates were incubated at 37°C in a humidified incubator with 5% CO₂. Then, 100 μL of 4T1 murine mammary cancer cells that have been pre-treated with 25 μg mL⁻¹ mitomycin C (MMC) (Sigma-Aldrich, USA) for 2 hr were added to the wells containing the freshly plated splenocytes. The plate was incubated at 37°C for 72 hr in a humidified 5% CO₂ incubator. After 72 hr, the splenocytes were harvested into 1.5 mL tubes and centrifuged (200 g for 10 min at 4°C). The culture supernatant from each sample was collected and stored at -80°C prior to quantification of interferon gamma (IFN-γ) and transforming growth factor β (TGF-β) using commercial READY SET GO! enzyme-linked immunosorbent assay (ELISA) kits as recommended by the manufacturer (eBiosciences, San Diego, Inc.).

**Statistical Analysis**

One-way analysis of variance (ANOVA) was performed using Statistical Package for Social Science (SPSS) version 16. The post-hoc Dunnett’s test was used to compare data of experimental group against control groups. All data points are expressed as the mean ± standard deviation (SD). A p-value of less than 0.05 (p<0.05) was considered to be statistically significant.

**RESULTS AND DISCUSSION**

There was a significant delay in tumour progression on days 21 and 28 in mice fed with γT3 compared to the vehicle-fed animals as shown by the tumour volume (Figure 1a) and tumour weight (Figure 1b) measurements. Histopathology findings on tumour sections from γT3-fed mice exhibited pleomorphic features with hyperchromatic nucleus
and the presence of necrosis (Figure 2) whilst the breast tissues sections from the vehicle-fed mice showed poorly differentiated tumour and marked metastasis. In addition, sections obtained from the vital organs of animals fed with γT3, such as the lung (Figure 3a), liver (Figure 3b) and kidney (Figure 3c) showed signs of minimal or delayed metastasis when compared to the vehicle-fed group. The lung tissue sections from both groups showed clusters of malignant (yellow circle) inflammatory (indicated with white circle) cells (Figure 3a). Sections of liver tissues from both groups also showed similar features (Figure 3b). However, sections of kidney tissue from both groups showed normal architecture with well-preserved glomerular and tubular structures (red circle) (Figure 3c). However, the tissue sections from the liver and lungs from the vehicle-fed mice showed presence of multiple metastasis foci, which appear to have developed rapidly (Table 1). These findings suggest that γT3 supplementation inhibited development and progression of BC as well as blocking the onset of metastasis. Previous studies have found that daily supplementation with TRF inhibited tumour growth and metastasis in this highly invasive syngeneic mouse model of BC (Abdul Hafid et al., 2013). Metastasis of tumour to distant organs has been associated with higher mortality (Ording et al., 2017) and in BC patients, tumour usually metastasises to their lungs (Medeiros and Allan, 2019) causing further complication. Inhibition of tumour growth following γT3 supplementation observed in this study is in agreement with our previous studies, where we had reported that daily supplementation of TRF inhibited tumour growth and metastasis in this syngeneic murine model of BC (Abdul Hafid et al., 2013; Abdul Hafid and Radhakrishnan, 2019; Selvaduray et al., 2010; Weng-Yew et al., 2009). Besides these evidence, recent studies also showed that γT3 significantly suppressed proliferation and invasion of prostate (Fontana et al., 2020), gastrointestinal (Zhang et al., 2018), colon (Aggarwal et al., 2019; Wada et al., 2017) and liver (Aggarwal et al., 2019; Sazli et al., 2015) cancer cells. The underlying mechanism of how γT3 suppresses tumour growth could be due to its anti-angiogenesis (Selvaduray et al., 2012), anti-proliferative (Nesaretnam et al., 1995) and anti-apoptotic (Srivastava and Gupta, 2006; Wu and Ng, 2010) properties; which are well-documented in the literature. However, in the present study, we intent to investigate the involvement of CD4, FoxP3, IL-24, IL-12Rβ2 in the γT3 mediated suppression of tumour growth and metastases.

Note: Tumour volume and weight were measured once every seven days (days 14, 21, 28 and 35) using a digital calliper. Tumour volume (mm³) was calculated using a formula that was previously described (Selvaduray et al., 2010). Tumour weight was measured by weighing the tumour at autopsy. Each data point represents mean (n=6) ± SD.

Figure 1. (a) Tumour volume, and (b) tumour weight.
### TABLE 1. KRUSKAL-WALLIS TEST OF LUNG AND LIVER METASTASIS

<table>
<thead>
<tr>
<th>Groups</th>
<th>Number of mice</th>
<th>Breast tumour differentiation</th>
<th>Metastasis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Lung</td>
</tr>
<tr>
<td>Vehicle</td>
<td>1</td>
<td>Poorly differentiated</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>Poorly differentiated</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>Poorly differentiated</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>Poorly differentiated</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>Moderately differentiated</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>Moderately differentiated</td>
<td>-</td>
</tr>
<tr>
<td>γT3</td>
<td>1</td>
<td>Necrosis</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>Moderately differentiated</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>Necrosis</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>Necrosis</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>Necrosis</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>6</td>
<td>Poorly differentiated</td>
<td>+</td>
</tr>
</tbody>
</table>

Note: The metastasis deposits in the lung and liver of vehicle and γT3 groups were compared as present (+) and absent (-). (p<0.05) for lung and liver.

---

Figure 2. Photomicrograph images (200X) of H&E stained sections of breast tumour tissues collected on day 21, 28 and 35 from animals fed with vehicle or γT3 (GT3) groups.
Figure 3. Photomicrograph images (200X) of H&E stained (a) lungs; (b) liver and (c) kidney sections collected at autopsy on day 0, 14, 21, 28 and 35 days following tumour induction in mice fed with vehicle or γT3 (GT3).

Note: Malignant cells indicated with yellow circle; inflammatory cells indicated with white circle.
CD4 is a cell surface glycoprotein found on surface of T-helper (Th) and Treg cells (Sambucci et al., 2019), which has high specificity to major histocompatibility complex class II (MHC II) proteins (Rzepecka et al., 2019). Activation of CD4+ T-cells can result in differentiation of sub-types that facilitate immune regulation through secretion of certain cytokines. A previous study by Huang et al. (2015) suggested that CD4+, along with CD8+ T-cells are dynamically involved in the immune response against BC, and that Th1 and CD8+ T-cells were dominant populations in tumour-infiltrating lymphocytes involved in the immunosurveillance in the early stage of BC development. However, in the late cancer stage, the number of CD4+ tumour-infiltrating lymphocytes increased significantly and the Treg and Th17 cells become the dominant populations, which contribute to the tumour promotion. The high number of CD4+ T-cells are most likely Treg cells, which also express this cell surface protein. High numbers of Treg cells are indicative of poor prognosis as these cells will create an immunosuppressive condition in the tumour microenvironment (Knochelmann et al., 2018; Ohue and Nishikawa, 2019). Hence, tumour will be able to progress due to the compromised cell-mediated immunity and reduction in the T-cell populations. BC patients and experimental animals induced with BC with advanced cancer frequently report dysfunctions in the immune system, which is evident from the reduced CD4+ to CD8+ ratio (CD4+:CD8+) and decreased T-cell proliferation possibly due to suppression by the Treg cells. In the present study, tumour tissue sections from γT3-fed mice showed higher number of CD4+ cells compared to tumour sections from vehicle-fed animals (Figure 4), suggestive of CD4+ associated anti-tumour immunity induced by γT3. However, it should be noted that Treg cells also express the CD4 glycoprotein on the cell surface (Yu et al., 2012).

The transcription factor FoxP3 is a key biomarker that can be used to identify Treg cells (Tanaka and Sakaguchi, 2019). Tumour tissue sections from vehicle fed mice showed presence of high number of FoxP3+ cells (Figure 4). High number of FoxP3+ cells in tumour microenvironment is associated with suppression of host immune response, which in turn can allow tumour progression to take place (Sasidharan and Elkord, 2018; Verma et al., 2019). However, tumour sections from γT3-fed mice showed marked reduction in the number of FoxP3+ cells (Figure 4). These findings suggest that γT3 supplementation can facilitate the host immune system to suppress tumour progression by limiting the immunosuppressive tumour microenvironments.

Tumour sections from γT3-fed animals showed higher expression of IL-24 and IL-12Rβ2 compared to sections from vehicle-fed mice (Figure 4), which supported presence of cell-mediated immunity. Interleukin-24 is cytokine linked with inhibition of tumour growth, anti-angiogenic and anti-metastasis activities (Panneerselvam et al., 2019; Zhang et al., 2019) whereby IL-12Rβ2 together with IFN-γ mediates differentiation of Th1 subset that plays crucial roles in cell-mediated immune responses, which include anticancer effects (Yamamoto et al., 1997). Statistical analysis using a semi-quantitative method confirmed that the difference observed on the expression of the four biomarkers (CD4, IL-24, IL-12Rβ2 and FoxP3) in tumour tissues section from vehicle- or γT3-fed mice were statistically significant (p<0.05) (Figure 5).

Figure 4. Photomicrograph images (200X) of breast tumour tissue sections from vehicle or γT3-fed mice stained with antibodies to murine (a) CD4, (b) FoxP3, (c) IL-12Rβ2 and (d) IL-24 biomarkers analysed using immunohistochemistry.
There was no significant difference ($p>0.05$) in the percentage of T-lymphocytes that express CD4$^+$ (TH) or CD8$^+$ (cytotoxic T-lymphocytes) as well as NK cells in peripheral blood at any of the time-points studied (Figure 6). There was a significant ($p<0.05$) increase in the Treg cell population in peripheral blood from vehicle-fed mice on day 28 and day 35 (Figure 6). The increase in the number of Tregs appears to correlate with bigger tumour load (Figure 1). These findings support the results from the immunohistochemistry analysis, which showed reduction in FoxP3$^+$ cells in the tumour tissue sections from γT3 fed mice. This provided more evidence to support that γT3 supplementation can modulate the host immune system in this syngeneic mouse model of BC.

However, it should be noted that γT3 supplementation did not completely stop tumour progression. One of the reasons for this observation may be because γT3 supplementation only started once the tumour is palpable. The syngeneic mouse model of BC used in this study is a highly tumourigenic model (Pulaski and Ostrand-Rosenberg, 2000). Hence, intervention with γT3 once tumour cells have been inoculated may give better results as this could trigger early activation of the host immune system to fight cancer. Another reason could be the low bioavailability of γT3 in the blood as previous studies have found that oral γT3 has low bioavailability due to low intestinal permeability (Abuasaal et al., 2012).

Culture supernatant from mitogen-stimulated splenocytes from vehicle- and γT3-fed mice showed significantly higher levels of IFN-γ (Figure 7) when compared to baseline (day 0). However, there was no significant difference between the two study groups. IFN-γ is the main regulator for TH1 immune response, which has a crucial role in controlling tumour growth (Alizadeh et al., 2021) whilst TGF-β is a major cytokine that supports activities of Treg cells (Mikami et al., 2020), which play an important role in immunosuppression i.e., allow tumour progression. The lack of significant difference in the production of these cytokines between the two study groups may be attributed to the delay in starting the γT3 intervention in this model. It is also possible that feeding γT3 for a short duration may not be sufficient to activate host immune response to secrete sufficient cytokines to suppress the cancer. The syngeneic mouse model of BC used in the present study is a highly invasive and spontaneously metastatic model. Hence, short term feeding of γT3 once tumour is palpable, may be insufficient to prepare the host immune system to control the tumour. Previous studies that have looked at effects of TRF supplementation in tumour-induced mice vaccinated with dendritic cell (DC) vaccines before (Abdul Hafid and Radhakrishnan, 2019; Abdul Hafid et al., 2013; Hafid et al., 2010) showed enhanced immune function and tumour suppression. However, it should be noted that in both these studies, the animals were supplemented with TRF for a longer period, which may have been sufficient to regulate the host immune system to fight against cancer.

To date, there has been only one study that investigated the effects of using TRF from palm oil in combination with tamoxifen in human BC patients (Nesaretnam et al., 2010). This study reported no association between adjuvant tocotrienol therapy and BC-specific survival in women with early BC, which has posed a setback to carry out more robust clinical trials in BC patients. More studies are needed to further evaluate the potential of using TRF as an anticancer agent in BC patients.
Note: Leucocytes were isolated from peripheral blood (vehicle or γT3 (GT3)-fed mice) by via cardiac puncture and stained with fluorochrome-labelled antibodies to various murine cell surface proteins and analysed using a flow-cytometer. Data was collected using the Cell Quest software provided by the manufacturer (Becton Dickinson, USA). In each acquisition, 10 000 cells were collected for data analysis. Results expressed as mean percentage ± SD of 6 mice per group. #p<0.05 when each group is compared against baseline (day 0); *p<0.05 when γT3 group is compared against vehicle group.

Figure 6. Percentage of (a) CD4+ T-cells, (b) CD8+ T-cells, (c) NK cells, and (d) Treg cell in peripheral blood analysed using a flow-cytometer.

Note: Culture supernatant was harvested after 72 hr and analysed for (a) IFN-g and (b) TGF-β using commercial ELISA kits. Results expressed as mean percentage ± SD of 6 mice per group. # Significantly different (p<0.05) from baseline (day 0).

Figure 7. Splenocytes obtained from spleens of mice fed with vehicle or γT3 (GT3)-fed mice cultured in the presence of mitomycin-C treated 4T1 cells.
CONCLUSION

This study shows that supplementing mice with 0.5 mg twice daily with γT3 showed promising results with regards to its anticancer effects based on the tumour growth and histopathology results. These findings suggest that γT3 provided via the oral route has reached the breast tumour tissues and was able to exert some anticancer effects in this mouse model. However, γT3 supplementation did not appear to have a significant effect on modulation of host immune system in this model.

ACKNOWLEDGEMENT

The authors would like to thank the International Medical University for providing the research facilities to carry out this study. This work was supported by a research grant from the Malaysian Palm Oil Board (MPOB) [PD167] and an FRGS grant (FRGS/1/2013/SG05/IMU/01/1). All the lab work was supported by a research grant from the Malaysian Palm Oil Board (MPOB) [PD167] and an FRGS grant supported by a research grant from the Malaysian Ministry of Health, Putrajaya. p. 1-100.

REFERENCES


Alizadeh, D; Kong, R A; Gholamin, S; Maker, M; Aftabizadeh, M; Yang, X; Pecoraro, J R; Jeppson, J D; Wang, D; Aguilar, B; Starr, R; Larmonier, C B; Larmonier, N; Chen, M-H; Wu, X; Ribas, A; Badie, B; Forman, S J and Brown, C E (2021). IFNg is critical for CAR T cell mediated myeloid activation and induction of endogenous immunity. Cancer Discov., 11(9): 2248-2265. DOI: 10.1158/2159-8290.CD-20-1661.


Huang, Y; Ma, C; Zhang, Q; Ye, J; Wang, F; Zhang, Y; Hunborg, P; Varvares, M A; Hoft, D F; Hsueh, E C and Peng, G (2015). CD4+ and CD8+ T cells have opposing roles in breast cancer progression and outcome. Oncotarget, 6(19): 17462-78. DOI: 10.18632/oncotarget.3958.


Panneerselvam, J; Srivastava, A; Mehta, M; Chen, A; Zhao, Y; D; Munshi, A and Ramesh, R (2019). IL-24 inhibits lung cancer growth by suppressing GLI1 and inducing DNA damage. Cancers, 11(12): 1879. DOI: 10.3390/cancers11121879.


Yu, N; Li, X; Song, W; Li, D; Yu, D; Zeng, X; Li, M; Leng, X and Li, X (2012). CD4+ CD25+ CD127 low/− T cells: A more specific Treg population in human peripheral blood. *Inflammation*, 35(6): 1773-1780. DOI: 10.1007/s10753-012-9496-8.
